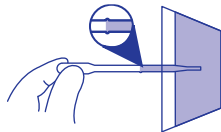


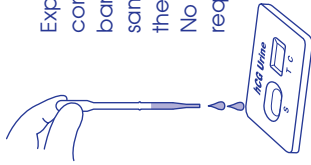


Squeeze the bulb of the pipette and draw up enough sample to fill the barrel to the line indicated on the pipette.



Do not overfill.

Expel **entire** contents of the barrel into the sample well of the test device. No drop counting required.



Read results at 3 minutes.

genzyme
Diagnostics

Rev. 3792-1, 03/06

OSOM Card Pregnancy Test

CLIA Complexity: Waived

INTENDED USE

The OSOM® Card Pregnancy Test is a rapid immunoassay for the qualitative detection of human chorionic gonadotropin (hCG) in urine as an aid in the early determination of pregnancy. This test is for professional use in physicians' offices and clinical laboratories.

SUMMARY AND EXPLANATION

Human chorionic gonadotropin (hCG) is a glycoprotein hormone secreted by the developing placenta. ⁽¹⁾ After fertilization, the concentration of hCG rapidly rises in both the urine and serum of pregnant women. The detection of hCG in these fluids is an excellent marker for confirming pregnancy. The OSOM Card Pregnancy Test is a rapid test which can detect the presence of hCG in urine. The test utilizes monoclonal and polyclonal antibodies to hCG.

PRINCIPLE OF THE TEST

OSOM Card Pregnancy Test is a solid phase, sandwich-format immunochromatographic assay for the qualitative detection of hCG. Urine is added to the sample well of the Test Device using the pipette provided. The sample migrates through reaction pads where hCG, if present in the sample, binds to a monoclonal anti-hCG dye conjugate. The sample then migrates across a membrane towards the results window, where the labeled hCG complex is captured at a test line region containing immobilized rabbit anti-hCG. Excess conjugate will flow past the test line region and be captured at a control line region containing an immobilized antibody directed against the anti-hCG dye conjugate (with or without hCG complexed to it).

The appearance of 2 black bands in the results window – one at "T: Test" and the other at "C: Control" – indicates the presence of hCG in the sample. If a detectable level of hCG is not present, only the control band will appear in the result window.

REAGENTS

- OSOM Card Pregnancy Tests Devices
- Membrane coated with rabbit polyclonal anti-alpha hCG
- Conjugate pad containing mouse monoclonal anti-beta hCG

WARNINGS AND PRECAUTIONS

- For in vitro diagnostic use. Federal law restricts this device to sale by or on the order of a physician.
- Do not use beyond the expiration date printed on the kit or foil pouch.
- The lot numbers may be different on the foil pouch and the kit.
- Use appropriate precautions for the collection, handling, and storage of specimens.
- Dispose of all used Test Devices, pipettes and specimens in suitable biohazardous waste containers.
- Test Devices are stable in the unopened foil pouches until the expiration date. Do not remove the Test Device from the pouch until needed.

STORAGE

Store OSOM Card Pregnancy Tests at room temperature, 15° to 30°C (59° to 86 °F), out of direct sunlight. Test Devices are stable until the expiration date printed on the kit or foil pouch. **DO NOT FREEZE.**

If the control band does not appear when running the test, the Test Cassette or kit may have been stored or handled improperly or the foil pouch may not have been intact.

SPECIMEN COLLECTION AND PREPARATION

No filtration or centrifugation of urine specimen is required for testing with the OSOM Card Pregnancy Test.

Urine specimens may be collected in any clean, dry, plastic or glass container. For early determination of pregnancy, the first morning specimen of urine is recommended.

since it usually contains the highest concentration of hCG. Urine specimens may be stored at room temperature 15° to 30°C (59° to 86°F) for up to 8 hours, or refrigerated at 2° to 8°C (35° to 46°F) for up to 72 hours.

QUALITY CONTROL Internal Quality Control

Several procedural controls are incorporated into each OSOM Card Pregnancy Test for routine quality checks. It is recommended that these procedural controls be documented for each sample as part of daily quality control.

The same labeled conjugate antibody results in the appearance of both the test and the control bands. The appearance of the control band in the results window is an internal positive procedural control which validates the following:

Test System: The appearance of the control band assures that the detection component of fluid was added to the sample well for capillary migration to occur. If the control band does not appear at the read time, the test is invalid. Appearance of the control band should be documented as part of the daily quality control.

Operator: The appearance of the control band indicates that an adequate volume of fluid was added to the sample well for capillary migration to occur. If the control band does not appear at the read time, the test is invalid. Appearance of the control band should be documented as part of the daily quality control.

The clearing of the background in the results area may be documented as a negative procedural control. It also serves as an additional capillary flow control. At the read time, the background should appear white to light gray and not interfere with the reading of the test. The test is invalid if the background fails to clear and obscures the observation of a distinct control band.

If the control band fails to appear with a repeat assay, do not report patient results. Contact Genzyme Diagnostics for technical service: Tel 617-252-7760 / 800-332-1042 (U.S. Customers only) / Fax 617-252-7759.

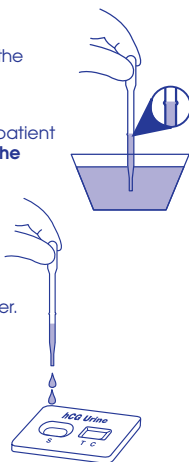
External Quality Control

Genzyme Diagnostics recommends that external hCG controls be run with each new lot, and with each new untrained operator. The OSOM hCG Urine Control (Catalog Number 134) is designed for this purpose. Quality Control requirements should be established in accordance with local, state and federal regulations or accreditation requirements.

TEST PROCEDURE

Patient specimens and control material must be brought to room temperature (15°-30°C; 59°-86°F) prior to testing.

- Remove the Test Device and the pipette from the pouch. Place the Device on a flat surface.
- Squeeze the bulb of the pipette and insert the barrel into the patient sample. Release the bulb and draw up enough sample to **fill the barrel to the line indicated on the pipette. Do not overfill.**
- Expel the entire contents of the barrel (135µL) into the sample well of the Test Device. No drop counting required.
- Discard the pipette in a suitable biohazardous waste container.
- Read results at 3 minutes
- Results are invalid after the stated read time. The use of a timer is recommended.**



MATERIALS PROVIDED

- OSOM Card Pregnancy Test Devices individually pouched, each containing a disposable pipette.
- Directional Insert

MATERIALS REQUIRED BUT NOT PROVIDED

- Clock or timer
- Sample collection cups or tubes
- Positive and Negative Controls (Genzyme Diagnostics recommends the OSOM hCG Urine Control (Catalog number 134)).

PROCEDURAL NOTES

- If specimen has been stored refrigerated, allow it to warm to room temperature before use.
- Several tests can be run at the same time. Use a new pipette with each test to avoid contamination errors.

LIMITATIONS IN HCG TESTING

- This assay is capable of detecting only whole molecule (intact) hCG. It cannot detect the presence of free hCG subunits. Therefore, this test should only be used for the qualitative detection of human chorionic gonadotropin in urine or serum for the early determination of pregnancy.
- For diagnostic purposes, hCG test results should always be used in conjunction with other methods and in the context of the patient's clinical information (e.g., medical history, symptoms, results of other tests, clinical impression, etc.). Ectopic pregnancy cannot be distinguished from normal pregnancy by hCG measurements alone.^(2,3)
- If the hCG level is inconsistent with, or unsupported by, clinical evidence, results should also be confirmed by an alternative hCG method. **Test results should be confirmed using a quantitative hCG assay prior to the performance of any critical medical procedure.**
- Interfering substances may falsely depress or falsely elevate results. These interfering substances may cause false results over the entire range of the assay, not just at low levels, and may indicate the presence of hCG when there is none. As with any immunochemical reaction, unknown interferences from medications or endogenous substances may affect results.
- Infrequently, hCG levels may appear consistently elevated and could be due to, but not limited to, the presence of the following:⁽⁴⁻⁷⁾
 - trophoblastic or nontrophoblastic neoplasms: abnormal physiological states that may falsely elevate hCG levels.^(8,9) This test should not be used in the diagnosis of these conditions.
 - hCG like substances

INTERPRETATION OF TEST RESULTS



Two separate black or gray bands – one at “T: Test” and the other at “C: Control” – are visible in the results window, indicating that the specimen contains detectable levels of hCG. While the intensity of the test band may vary with different specimens, the appearance of 2 distinct bands should be interpreted as a positive result.



If no band appears at “T” and a black or gray band is visible at the “C: Control” position the test can be considered negative, indicating that a detectable level of hCG is not present.



If no band appears at the “C: Control” position, the test is invalid. The test is also invalid if incomplete or beaded bands appear at either the “T: Test” or “C: Control.” The test should be repeated using another Test Device.

Note: The test is valid if the control line appears by the stated read time regardless of whether the sample has migrated all the way to the end of the sample window.

- Because of the high degree of sensitivity of the assay, specimens tested as positive during the initial days after conception may later be negative due to natural termination of the pregnancy. Overall, natural termination occurs in 22% of clinically unrecognized pregnancies and 31% of other pregnancies.⁽¹⁰⁾ In the presence of weakly positive results, it is good laboratory practice to sample and test again after 48 hours.
- If the test band appears very faint, it is recommended that a new sample be collected 48 hours later and tested using another OSOM Card Pregnancy Test Device.
- Dilute urine specimens may not have representative levels of hCG.
- Detection of very low levels of hCG does not necessarily indicate pregnancy⁽⁴⁾ as low levels of hCG can occur in apparently healthy, nonpregnant subjects.^(11,12) Additionally, post-menopausal specimens may elicit weak positive results due to low hCG levels unrelated to pregnancy. In a normal pregnancy, hCG values double approximately every 48 hours.⁽¹³⁾ Patients with very low levels of hCG should be sampled and tested again after 48 hours, or tested with an alternative method.
- Some antipsychotic agents/drugs are known to cause false positive results in pregnancy tests.⁽¹⁴⁾

EXPECTED VALUES

hCG is not normally detected in the urine and serum specimens of healthy men and non-pregnant women. In normal pregnancy, 20 mIU/mL hCG is reported to be present in both urine and serum 2 to 3 days before the first missed menstrual period.^(15,16) The levels of hCG continue to increase up to 200,000 mIU/mL at the end of the first trimester.

Agreement

Urine specimens from 634 individuals were evaluated with the OSOM Card Pregnancy Test and the QuickVue[®]+ One-Step hCG-Combo Test. Samples were from patients seeking confirmation of pregnancy. The two assays were in agreement on 629 of the 634 samples. A radioimmunoassay (DPC Coat-A-Count[®] hCG IRMA Kit) was used to quantify the five discrepant results. Three of the discrepant samples were found to have an hCG concentration greater than 0 but less than 20 mIU/mL, the stated analytical sensitivity of both assays, and thus were removed from the analysis. One sample contained 0 mIU hCG/mL according to the IRMA and was scored negative by the OSOM test but positive by QuickVue+. The remaining sample contained >500 mIU hCG/mL according to the IRMA and was scored positive by the OSOM test but negative by QuickVue+.

Thus in this study, the OSOM hCG urine procedure had greater than 99% agreement with the comparative test methods in the 435 specimens testing negative and the 196 specimens testing positive.

Comparative Methods
(QuickVue+ Test and IRMA)

OSOM Card Pregnancy Test

	+	-
+	196	0
-	0	435

Agreement on Positive Samples: >99%
Agreement on Negative Samples: >99%
Total Agreement: >99%

Physician's Office Laboratory (POL) and Laboratory Study

A proficiency panel was prepared to allow for the evaluation of the urine testing format at three physician's office and a clinical laboratory. A total of 40 samples were tested at each site. Purified hCG was spiked into an artificial urine matrix. Each panel contained negative, low positive, moderate positive and high positive samples. Each panel was tested at each site over the course of three distinct runs. 100% of the positive and negative results obtained by the POL operators were in agreement with the expected values and with the results obtained by the clinical laboratory operators.

PERFORMANCE CHARACTERISTICS

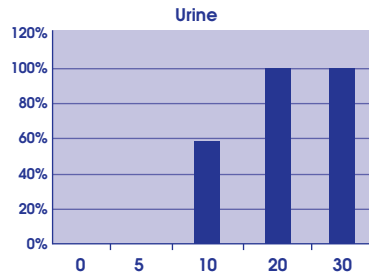
Sensitivity

The OSOM Card Pregnancy Test will detect hCG in urine specimens with concentrations of 20 mIU/mL or more (calibrated against WHO 3rd IS 75/537). Specimens containing 1,000,000 mIU/mL (spiked with purified hCG) will also give positive results.

- The expected sensitivity of urine samples at a read time of 3 minutes is 20 mIU/mL

Note: Samples containing minute quantities of hCG (below 10 mIU/mL) may develop faint test bands.

% of samples containing varying amounts of hCG interpreted as positive



Cross Reactivity

The addition of luteinizing hormone (300 mIU/mL of LH), follicle stimulating hormone (1000 mIU/mL of FSH), or thyroid stimulating hormone (1000 µIU/mL of TSH) to negative urine specimens gives negative results in the OSOM Card Pregnancy Test.

Interfering Substances

The following substances were added to urine specimens containing 0 or 20 mIU/mL hCG. The substances at the concentrations listed below were not found to affect the performance of the test.

Acetaminophen	20 mg/dL	Gentisic acid	20 mg/dL
Acetoacetic acid	2000 mg/dL	Glucose	2000 mg/dL
Acetyl salicylic acid	20 mg/dL	Hemoglobin	250 mg/dL
Amitriptyline	100 mg/dL	Human albumin	2000 mg/dL
Amphetamines	10 µg/mL	Ibuprofen	40 mg/dL
Ascorbic acid	20 mg/dL	Imipramine	100 mg/dL
Atropine	20 mg/dL	Lithium	3.5 mg/dL
Benzoylcogonine	10 mg/dL	Mesoridazine	1mg/dL
Bilirubin	2 mg/dL	Methadone	10 mg/dL
Caffeine	20 mg/dL	Morphine	6 µg/mL
Cannabinol	10 mg/dL	Nortriptyline	100 mg/dL
Chlorpromazine	5 mg/dL	Phenobarbital	15 mg/dL
Codeine	10 mg/dL	Phenylpropanolamine	20 mg/dL
Desipramine	20 mg/dL	Pregnanediol	1500 µg/dL
Diazepam	2 mg/dL	Progesterone	40 ng/mL
Ephedrine	20 mg/dL	Proteins	2000 mg/dL
Estradiol	25 ng/mL	Salicylic acid	20 mg/dL
Estriol	1 mg/dL	Tetracycline	20 mg/dL
Hydroxybutyrate	2000 mg/dL	Thioridazine	2 mg/dL
Ethanol	200 mg/dL		

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ASSISTANCE

For technical assistance, call Genzyme Diagnostics technical service at 800-332-1042.

REORDER

No. 102 (25 tests)

TRADEMARK

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